| 1 | Control of African lovegrass by flupropanate in a flora conservation context | | | |
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| 10 | | | | |
| 11 | Article type: Research paper | | | |
| 12 | Word Count: Abstract - 250 Main text (incl. references) - 3332 | | | |
| 13 | | | | |
| 14 | Number of Tables: 1 | | | |
| 15 | Number of figures: 4 | | | |
| 16 | | | | |

17 Abstract

Context. Managing widespread invasive plants to support biodiversity conservation is a 18 significant challenge in many ecosystems, and requires weed control methods that have 19 acceptable impacts on co-occurring native species. African lovegrass (Eragrostis curvula) is a 20 perennial grass that has become invasive in many regions globally. There is a lack of effective 21 control options, particularly in diverse native vegetation where application of broad-spectrum 22 23 herbicide has risks of unacceptable off-target impacts. Aims. We tested the effectiveness of flupropanate (sodium 2,2,3,3-tetra-fluoropropionate) in controlling African lovegrass in a 24 25 conservation context in Mediterranean-climate south-western Australia, testing two application rates and measuring target and off-target impact. Methods. Cover of plant species 26 and their condition (alive or dead) was measured in replicate plots in an 'before-after-control-27 impact' design. A small sample of endangered Grevillea curviloba individuals were 28 deliberately treated with flupropanate. Key results. Flupropanate significantly reduced 29 African lovegrass cover, with greater reduction at the higher application rate. No significant 30 off-target effects could be detected at a community (excluding African lovegrass) or plant 31 32 functional group level nor in the deliberate exposure of Grevillea curviloba. However, there was a suggestion of increased mortality in one native species incidentally exposed to 33 flupropanate. Conclusions. Land managers in south-western Australia can have confidence 34 that flupropanate will be effective in controlling African lovegrass in conservation contexts, 35 36 and specifically where infestations co-occur with G. curviloba. Implications. Evidence of off-target impacts emerging from this and other studies indicates that robust testing of the 37 susceptibility of conservation-listed flora to flupropanate should precede any application in 38 39 their habitat.

41 Keywords: grass weeds; selective herbicide; off-target impact; threatened flora; weed
42 management

43

44 Introduction

Weed invasions have substantial impacts on biodiversity (Pyšek et al. 2012). Effective control
methods are thus needed to address weed threats. In a biodiversity conservation context for
the management of widespread weeds, acceptably low off-target impacts of weed control
methods are required for management investment to achieve the desired conservation
outcomes (Farmillo and Moxham 2023).

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African lovegrass (Eragrostis curvula (Schrad.) Nees), a long-lived tussock grass with a 51 52 native range of southern Africa, has become a significant weed globally including in Australia and North America (Roberts et al. 2020). It has been introduced widely through seed 53 contamination and deliberately as a pasture species and for soil stabilization, but has spread 54 further to become a weed in both agricultural and environmental settings (Firn 2009; van 55 Klinken et al. 2017). As a perennial grass using the C4 photosynthetic pathway, African 56 lovegrass grows actively through the warmer months of the year. African lovegrass has the 57 capacity to resprout strongly from the base of tussocks following fire and other disturbances, 58 59 and its spread is primarily via dispersal of seed by machinery, wind and externally on 60 animals.

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The most common method for control of African lovegrass in conservation settings is
application of the non-selective herbicide glyphosate, although mechanical removal can be
effective for low-density infestations, and preventing spread is also an important African
lovegrass management strategy (Firn 2009; Blakely et al. 2022). African lovegrass control in

conservation settings is challenging as there are risks of off-target impacts in the use of 66 broad-spectrum herbicides. As an alternative to glyphosate, flupropanate (sodium 2,2,3,3-67 tetra-fluoropropionate; marketed under several product names) has been demonstrated to be 68 effective for African lovegrass control in some eastern Australian agricultural and 69 70 environmental situations (Campbell and Nicol 1998). While flupropanate may have potentially less uniform impact on co-occurring vegetation than glyphosate (Campbell et al. 71 72 2002), off-target impacts on vegetation ranging from tree seedlings to desirable pasture species have been recorded (McLaren et al. 2008; Lusk et al. 2017; Blakely et al. 2022). 73 74 Flupropanate has a low contact activity, so it is mainly absorbed into the soil and taken up by plant roots after rain post-application. Flupropanate effectiveness is thought to be dependent 75 on both temperature (acting faster with warmer conditions) and rainfall (requiring wet 76 77 conditions for activation) (Lusk et al. 2017). Some soil residual effects are typical but may be short-lived (Bourdôt et al. 2017). 78

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African lovegrass is regarded as among the highest impact grass environmental weeds in 80 Australia (van Klinken et al. 2017). African lovegrass is competitively superior to native 81 grasses under a range of environmental conditions and higher African lovegrass biomass is 82 associated with lower richness of co-occurring species (Firn et al. 2010; Firn et al. 2017). In a 83 weed prioritization for the Department of Biodiversity, Conservation and Attractions' 84 85 (DBCA) Swan Region in south-western Australia, African lovegrass was ranked with high ecological impact and rapid invasiveness (https://www.dbca.wa.gov.au/management/threat-86 management/weeds). The Swan Coastal Plain, which forms part of the Southwest Australian 87 88 Floristic Region biodiversity hotspot, supports an exceptional concentration of threatened flora, particularly on the relatively older (late Pliocene-middle Pleistocene) Bassendean 89 sands and Guildford sediments (Gosper et al. 2021a, 2022) on the eastern side of the plain. 90

| 91 | Levels of weed invasion are relatively high on the comparatively fertile Guildford sediment | | | | |
|-----|---|--|--|--|--|
| 92 | (Gosper et al. 2021b), with African lovegrass particularly problematic on the heavier soils | | | | |
| 93 | typical of this formation. African lovegrass invasion impacts a range of threatened flora | | | | |
| 94 | including Grevillea curviloba (Western Australian conservation status of endangered; Figure | | | | |
| 95 | 1), Darwinia foetida (endangered), Synaphea sp. Fairbridge Farm (D. Papenfus 696) | | | | |
| 96 | (critically endangered), Synaphea sp. Pinjarra (R. Davis 6578) (critically endangered) and | | | | |
| 97 | Synaphea sp. Pinjarra Plain (A.S. George 17182) (endangered), and threatened ecological | | | | |
| 98 | communities, such as Corymbia calophylla – Kingia australis woodlands on heavy soils | | | | |
| 99 | (critically endangered) and Corymbia calophylla – Xanthorrhoea preissii woodlands and | | | | |
| 100 | shrublands (endangered). | | | | |
| 101 | | | | | |
| 102 | 2 The aims of this study were to build on previous research by testing the efficacy of | | | | |
| 103 | flupropanate to control African lovegrass in a different climatic and land use context, | | | | |
| 104 | specifically to: | | | | |
| 105 | • Determine if flupropanate remains effective in controlling African lovegrass in a | | | | |
| 106 | strongly Mediterranean climate, where herbicide effectiveness could plausibly be | | | | |
| 107 | reduced by lower water availability over the warmer months of optimal African | | | | |
| 108 | lovegrass growth. | | | | |
| 109 | • Document any off-target effects on co-occurring native flora when flupropanate is | | | | |
| 110 | applied to control African lovegrass for conservation outcomes. | | | | |
| 111 | | | | | |
| 112 | Materials and methods | | | | |
| 113 | Study site and treatments | | | | |
| 114 | An area near Bullsbrook, north-east of Perth in southwest Western Australia, was chosen for | | | | |
| 115 | the study, having an extensive African lovegrass infestation adjoining and in similar habitat | | | | |

(soils, remnant vegetation) to key threatened flora in which African lovegrass control is
required. The site, in a narrow (less than 100m wide) and linear rail easement vested in the
Perth Transport Authority and the City of Swan, had been historically cleared of large trees,
but retained scattered mature native shrubs, over a mixed ground layer of dense African
lovegrass, invasive perennial veldt grass (*Ehrharta calycina*), invasive annual grasses, and
native and invasive herbs, geophytes and graminoids.

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The experiment was set up as a repeated measures 'before-after-control-impact' design. Three 123 124 sets of replicate 10×10 m plots were established and sampled in 2019 prior to application of each of three treatments; a no herbicide control, and a lower (1.5mL L^{-1}) and higher (3.0 mL)125 L^{-1}) flupropanate application rate. Plot corners were marked with steel droppers and recorded 126 with GPS. Liquid herbicide (mixed with dye) was applied by a ground-based operator with 127 backpack-based spray equipment as it would be in a management scenario, by targeted spot 128 spray to runoff of live African lovegrass plants. Due to high African lovegrass density, 129 exposure of co-occurring native and other invasive flora to flupropanate was inevitable and 130 because flupropanate uptake is primarily via the roots, even larger shrubs and small trees may 131 have been exposed. Treatments were applied in late spring (November) 2019, when African 132 lovegrass was actively growing (Roberts et al. 2021), with follow-up spot spraying applied as 133 required in November 2020. 134

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136 Vegetation measurement

At each of the nine plots, species presence and condition (whether the whole plant was alive or dead) were recorded at 200 point intercepts with an 12mm pole distributed 0.25m apart in a grid pattern along five equally-spaced transects across the plot. This technique provided an objective measure of species abundance/cover as the number of intercepts with the species

present, either alive and/or dead, out of 200. Any additional species present in the plot but not 141 intercepted were also recorded and allocated a nominal cover value of 0.5. Vegetation 142 measurements were completed in 2019 (pre-treatment), 2020 (~1 year after the first treatment 143 in 2019) and 2021 (~1 year after second treatment in 2020) immediately prior to herbicide 144 application. Consideration was given to normal modes of seasonal growth, so annual species, 145 or annually active species such as geophytes, were regarded as 'alive' in vegetation 146 147 measurements regardless of whether they had begun to senesce, due to senescing being the expected condition in late spring. Annual or annually active species in the post-treatment 148 149 samples would have germinated or sprouted after the previous herbicide treatment, so their detection indicates that they were able to grow after treatment of their habitat, with the effect 150 of treatments being assessed through changes in the number of intercepts. In contrast, any 151 susceptibility of above-ground perennial species can be assessed through both changes in the 152 quantity of live and dead intercepts. 153

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155 *Statistical analysis*

Cover of plant species was aggregated into functional groups on the basis of growth form
(annual, perennial groundcover (grass, herb and graminoid), and woody shrub) and status as
native or invasive, using the information from Western Australian Herbarium (1988-2024)
(Supplementary Material S1). Functional groupings did not include cover of African
lovegrass.

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162 Summed plot live vegetation intercepts (cover) for African lovegrass, invasive annuals,

163 invasive perennial groundcovers, native perennial groundcovers and native woody shrubs

164 were analyzed using the repeated measures ANOVA module in Statistica 7.1

165 (https://docs.tibco.com/products/tibco-statistica-14-0-0), with a fixed factor of treatment and

the fixed repeated measure of sample year (2019, 2020, 2021). Cover of African lovegrass
was log₁₀ transformed.

168

Ordination was used to explore changes in species composition over time and with 169 treatments. The cover data were filtered to live touches only, singletons were removed, and 170 separate data files prepared including and excluding cover of African lovegrass. Data were 171 172 square-root transformed and non-metric multidimensional scaling applied using the Bray-Curtis dissimilarity metric in PRIMER analysis software (version 6.1.11, https://www.primer-173 174 e.com/software). PERMANOVA, using a design a fixed factor of treatment, a random factor of site nested in treatment, and the fixed repeated measure of sample year (Anderson et al. 175 2008), was used to test for differences among herbicide treatments and time. 176

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178 Targeted exposure of Grevillea curviloba

Grevillea curviloba is one of the threatened flora for which an effective non-broad spectrum 179 herbicide for African lovegrass control would be highly valuable. To test the effect of 180 exposure to flupropanate of G. curviloba, three potentially 'sacrificial' plants were sprayed 181 with the high dose 3.0mL L⁻¹ liquid flupropanate treatment, with three unsprayed controls. 182 The 'sacrificial' plants were growing along a road verge and are periodically slashed (Figure 183 1), and hence are artificially maintained in a prostrate growth form. Sample plants were 184 marked by a metal tag attached to a peg inserted into the ground near the base of the plant, 185 and canopy dimensions (north-south and east-west) and condition were assessed, prior to 186 spraying (November 2019) and at 1, 6, 12 and 24 months afterwards. 187 188

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189 **Results**

Application of flupropanate resulted in a significant reduction in African lovegrass cover
(Figure 2), demonstrated by a significant time × treatment interaction. The higher application
rate resulted in a greater decline in African lovegrass cover in the first year post-treatment,
and although a second application further reduced African lovegrass cover at both herbicide
rates, the difference between application rates was maintained.

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196 No effects of flupropanate treatment were detected for any of the invasive annual, invasive perennial groundcover, native perennial groundcover or native shrub functional groups (Table 197 198 1). For invasive annuals, there was a significant effect of Year, with greater cover in the two post-treatment years. While there were insufficient intercept data and a lack of replication 199 across the sample plots for robust statistical analysis at the species level for native flora, there 200 201 was a suggestion of an adverse impact to flupropanate in Desmocladus virgatus (Figure 3). There was both an increase in dead intercepts and a decline in live intercepts in the first year 202 post-treatment in all flupropanate-treated plots where this species was recorded which was 203 not apparent in the control plot with D. virgatus. Live cover of D. virgatus recovered 204 somewhat following the second year of treatment. 205

206

Flupropanate application had a significant effect on plant community composition when all 207 species were included, as evidenced by a significant treatment × time interaction in 208 209 PERMANOVA (Figure 4a). In ordination, two of the three control plots remained highly similar in composition across all years, while all flupropanate-treated plots showed a 210 consistent directional shift in location in multivariate space over time associated with lower 211 African lovegrass cover. In contrast, using the same data except for excluding cover of the 212 target weed African lovegrass, there were no significant effects of flupropanate application on 213 community composition (Figure 4b). Plots did vary in composition (significant site nested in 214

treatment effect), and over time, reflected in the associations of the perennial native shrubs *Acacia saligna* and *Xanthorrhoea* sp. and invasive annuals *Bromus diandrus* and *Avena barbata* with the location of sites in multivariate space respectively.
Intentional exposure of three *G. curviloba* individuals to flupropanate did not result in any

mortality over the period of sampling, nor any obvious change in plant condition. Similarly,
the three control (no herbicide) individuals also survived.

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223 Discussion

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African lovegrass was effectively controlled by flupropanate, as has been found previously 225 226 (Campbell and Nicol 1998), indicating that the strongly Mediterranean climate of the study area did not affect the herbicide's effectiveness. Similarly to Bourdôt et al. (2017) with 227 *Nassella trichotoma*, higher levels of weed control were achieved at the higher (3.0mL L^{-1}) 228 flupropanate application rate. Land managers in south-western Australia can therefore have 229 confidence that flupropanate will be effective in controlling African lovegrass in conservation 230 contexts, and specifically where infestations co-occur with the endangered G. curviloba, 231 which was not detectably impacted by deliberate exposure. Flupropanate resistance in African 232 lovegrass has been recorded in NSW (Powells 2022), emphasizing the value of an integrated 233 234 African lovegrass control program.

235

As co-occurring flora species were in general either too infrequently encountered or did not occur in most plots, robust statistical analysis of the effects of flupropanate at the species level was not possible, so species were aggregated into functional groups and considered at the community level. There were no significant effects of flupropanate application on non-

target groups and at the community level (with African lovegrass excluded), noting these 240 findings refer to the effects of incidental exposure of non-target species to herbicide rather 241 than targeted exposure. The greater cover of invasive annuals in post-treatment years is 242 potentially explained by annual weeds increasing in abundance following African lovegrass 243 control, as has been found elsewhere following invasive perennial grass control with 244 flupropanate (Lusk et al. 2017). However, the increased invasive annual cover in post-245 246 treatment years was also found in control plots (no time × treatment interaction), suggesting variable seasonal conditions may be a more parsimonious explanation. 247

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However, there was an indication of increased mortality in D. virgatus with flupropanate 249 application, although more robust testing is required for confirmation. D. virgatus 250 251 (Restionaceae) is a rhizomatous, tufted, perennial herb, and as a low-growing plant would readily have been exposed to herbicide applied to co-occurring African lovegrass. As 252 flupropanate application was directed towards observable live African lovegrass individuals, 253 the partial recovery of D. virgatus following the second year of treatment could be explained 254 by the high levels of African lovegrass mortality after the initial treatment leading to lower 255 spatial coverage of flupropanate in the second treatment year, and hence a lower probability 256 of D. virgatus being sprayed. A potential explanation of the lower cover of dead D. virgatus 257 after the second treatment could be that dead D. virgatus material from the first treatment 258 259 year may have decayed to the point of being incorporated into leaf litter. As some off-target effects of flupropanate were likely in this study and have been shown elsewhere (McLaren et 260 al. 2008; Lusk et al. 2017), robust testing of conservation-listed flora's susceptibility to the 261 262 herbicide should precede any application in their habitat.

DBCA is further testing the effectiveness of flupropanate for control of other weed species of 264 the Swan Coastal Plain and monitoring for off-target impacts on native flora in intact 265 occurrences of two critically endangered threatened ecological communities, Banksia 266 attenuata and/or Eucalyptus marginata woodlands on the eastern side of the Swan Coastal 267 Plain and Corymbia calophylla – Kingia australis woodlands on heavy soils. The 268 investigation of the trajectory of vegetation composition after weed control by flupropanate in 269 270 native vegetation in good or better condition will be valuable, as in pasture settings replacement of the target weed by other weeds and bare ground has been the (undesirable) 271 272 outcome (Lusk et al. 2017). In this study, insufficient time had elapsed after treatment to discern any patterns of vegetation change, with the dead thatch of African lovegrass 273 remaining the dominant cover on treated plots. 274

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Supplementary Material - Taxa recorded from plots and their classification into functional
groups based on life form and status as native or invasive

278 Acknowledgements - We thank Anne Harris and David Mitchell for assistance with

279 conceptualization and fieldwork, and Sandra Williamson, Ebony Skey, Megan Young, John

280 Dagnall, Amy Gaunt, Simon Caunter and Alan Jenkins for assistance with fieldwork. John

281 Morrell from Arc Infrastructure and Brad Thompson from the City of Swan facilitated access

to the study sites.

Declaration of funding - This project received no specific funding.

284 Author contributions – All authors conceptualized and designed the study, undertook data

collection and revised the manuscript. CRG led data analysis and writing.

Conflict of interests - The authors declare no conflicts of interest.

- 287 **Data availability -** Raw data will be made publicly available through the Department of
- 288 Biodiversity, Conservation and Attractions' Data Catalogue upon article acceptance.

289 <u>https://data.bio.wa.gov.au/</u>

- **Permits** The work was conducted under section 40 authorization TFL 106-1920.
- 291

292 References

- Anderson, M.J., Gorley, R.N. and Clarke, K.R. (2008) *PERMANOVA+ for PRIMER: guide to software and statistical methods*. PRIMER-E, Plymouth.
- Blakely, S., Vitelli M., Tully M, Johnson, A.-M. and Colley J (2022) The advancing front of
- invasive lovegrasses across Australia's rangelands. *Proceedings of the 22nd Australasian*

297 Weeds Conference Adelaide September 2022 (Ed. R. Melland, C. Brodie, J. Emms, L.

- Feuerherdt, S. Ivory and S. Potter), pp. 150-153. Weed Management Society of SouthAustralia, Adelaide.
- 300 Bourdôt, G., Jackman, S. and Saville, D. (2017) Plant mortality and seedling recruitment
- 301 responses to flupropanate in grassland populations of *Nassella trichotoma*. *New Zealand*
- 302 *Plant Protection* 70, 160–164. DOI: <u>https://doi.org/10.30843/nzpp.2017.70.42</u>.
- 303 Campbell, M.H. and Nicol, H.I. (1998) Effects of wiping herbicides on serrated tussock
- 304 (*Nassella trichotoma* (Nees) Arech.) and African lovegrass (*Eragrostis curvula* (Shrad.)
- Nees). *Plant Protection Quarterly* 13, 36-38.
- 306 Campbell, M.H., Vere, D.T. and Nicol, H.I. (2002) Long-term control of serrated tussock
- 307 (*Nassella trichotoma* (Nees) Arech.) by applying flupropanate at three-year or 10-year
 308 intervals. *Plant Protection Quarterly* 17, 58-63.
- 309 Farmilo, B.J. and Moxham, C. (2023) The effectiveness of weed control in a threatened plant

community: A grassland case study. *Ecological Engineering* 193, 107017.

311 https://doi.org/10.1016/j.ecoleng.2023.107017

- Firn, J. (2009) African lovegrass in Australia: a valuable pasture species or embarrassing
 invader? *Tropical Grasslands* 43, 86-97.
- Firn, J., MacDougall, A.S., Schmidt, S., & Buckley, Y.M. (2010) Early emergence and
- resource availability can competitively favour natives over a functionally similar invader.
- 316 *Oecologia* 163, 775–784.
- Firn, J., Ladouceur, E., Dorrough, J. (2017) Integrating local knowledge and research to
- refine the management of an invasive non-native grass in critically endangered grassy
- 319 woodlands. *Journal of Applied Ecology* 55, 321–330. <u>https://doi.org/10.1111/1365-</u>
- <u>2664.12928</u>
- 321 Gosper, C.R., Coates, D.J., Hopper, S.D., Byrne, M. and Yates, C.J. (2021a) The role of
- 322 landscape history in the distribution and conservation of threatened flora in the Southwest
- Australian Floristic Region. *Biological Journal of the Linnean Society* 133, 394-410.
- 324 <u>https://doi.org/10.1093/biolinnean/blaa141</u>
- 325 Gosper, C.R., Kinloch, J., Coates, D.J., Byrne, M., Pitt, G. and Yates, C.J. (2021b)
- 326 Differential exposure and susceptibility to threats based on evolutionary history: how
- 327 OCBIL theory informs flora conservation. *Biological Journal of the Linnean Society* 133,
- 328 373-393. <u>https://doi.org/10.1093/biolinnean/blaa170</u>
- 329 Gosper, C.R., Percy-Bower, J.M., Byrne, M., Llorens, T.M. and Yates, C.J. (2022)
- 330 Distribution, biogeography and characteristics of the threatened and data-deficient flora in
- the Southwest Australian Floristic Region. *Diversity* 14, 493.
- 332 <u>https://doi.org/10.3390/d14060493</u>
- 333 McLaren, D.A., Snell, K. and Butler, K. (2008) An assessment of native tree susceptibility to
- the simulated aerial application of the herbicide flupropanate, for management of exotic
- 335 unpalatable grasses. Proceedings of the 16th Australian Weeds Conference (Ed. R.D. van

- Klinken, V.A. Osten, F.D. Panetta and J.C. Scanlan), pp. 323-325. Queensland Weeds
 Society, Brisbane.
- 338 Lusk, C. S., Hurrell, G. A., Saville, D. J., & Bourdôt, G. W. (2017). Changes in plant species
- composition after flupropanate application for nassella tussock control, in Canterbury hill-
- 340 country pastures. *New Zealand Journal of Agricultural Research* 60, 263–276.
- 341 https://doi.org/10.1080/00288233.2017.1321556
- Powells, J. (2022). Herbicide resistance in perennial pasture systems the horse has bolted.
- 343 *Proceedings of the 22nd Australasian Weeds Conference Adelaide September 2022* (Ed. R.
- Melland, C. Brodie, J. Emms, L. Feuerherdt, S. Ivory and S. Potter), pp. 17-20. Weed
- 345 Management Society of South Australia, Adelaide.
- 346 Pyšek, P., Jarošík, V., Hulme, P.E., Pergl, J., Hejda, M., Schaffner, U. and Vilà, M. (2012). A
- 347 global assessment of invasive plant impacts on resident species, communities and
- 348 ecosystems: the interaction of impact measures, invading species' traits and environment.
- 349 *Global Change Biology* 18, 1725-1737. <u>https://doi.org/10.1111/j.1365-2486.2011.02636.x</u>
- 350 Roberts J., Florentine S., van Etten E., and Turville C. (2021) Germination biology,
- distribution and control of the invasive species *Eragrostis curvula* [Schard. Nees] (African
- 352 Lovegrass): A global synthesis of current and future management challenges. *Weed*
- 353 *Research* 61, 154–163. <u>https://doi.org/10.1111/wre.12474</u>
- 354 Van Klinken, R.D. and Friedel, H.H. (2017) Unassisted invasions: understanding and
- responding to Australia's high-impact environmental grass weeds. *Australian Journal of*
- 356 *Botany* 65, 678–690. <u>https://doi.org/10.1071/BT17152</u>
- 357 Western Australian Herbarium (1998–2024). Florabase—the Western Australian flora.
- 358 Department of Biodiversity, Conservation and Attractions. Online:
- 359 <u>https://florabase.dbca.wa.gov.au/</u> (Accessed 12 November 2024).

Figure legends

Figure 1. (a) African lovegrass (*Eragrostis curvula*) invasion impacting a population of the
endangered *Grevillea curviloba*; (b) habitat of roadside *G. curviloba* plants that were
regularly slashed and which were tested for tolerance to flupropanate. Photos: Carl Gosper,
Julia Cullity.

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Figure 2. Effects treatment with flupropanate (control - untreated; low rate -1.5mL L⁻¹; high rate -3.0mL L⁻¹) on cover of live African lovegrass (*Eragrostis curvula*). ANOVA results (in text box) are based on log₁₀ transformed cover values (the number of point intercepts out of 200 with the species per plot), which were converted to proportional cover for graphical presentation. *** P<0.001, ** P<0.01, * P<0.05.

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Figure 3. Effects of treatment with flupropanate (control – untreated; low rate - $1.5mL L^{-1}$; high rate – $3.0mL L^{-1}$) on cover of *Desmocladus virgatus*, showing intercepts with live *Desmocladus* plants (green lines and symbols) and dead *Desmocladus* plants (cyan lines and symbols). *Desmocladus virgatus* was present in 4 of the 9 plots (two in the herbicide low treatment, with these data showing means \pm SE) and occurred so infrequently that robust statistical analysis was not possible.

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Figure 4. Effects treatment with flupropanate (\checkmark control - untreated; \blacktriangle low rate – 1.5mL L⁻¹; high rate – 3.0mL L⁻¹) on community composition based on plant cover (A) including all species; and (B) excluding cover of African lovegrass (*Eragrostis curvula*). Ordinations are non-metric multidimensional scaling with (A) stress 0.15, vector showing African lovegrass; (B) stress 0.13, vectors showing species with a Pearson's correlation > 0.8. PERMANOVA results (in text boxes) give pseudo-F values for effects of Treatment (Tr), sample Year (Yr),

- Site nested in Treatment (S(Tr)) and the interactions of Tr \times Yr. *** P<0.001, ** P<0.01, *
- 386 P<0.05.

Table 1. Statistical analysis results of the effect of application of flupropanate at two
applications rates on the cover of functional groups of plants (excluding African lovegrass),
showing ANOVA F values. Treatment (Tr) is the effect of levels of flupropanate, Year (Yr) is
the effect of sample year (pre-treatment, year 1 and 2 post-treatment), and Tr × Yr is the
interaction between these effects.

392

| | Tr | Yr | $Tr \times Yr$ | |
|---------------------------------|------|-------|----------------|--|
| df | 2,6 | 2,12 | 4,12 | |
| Functional group | | | | |
| Invasive annuals | 0.07 | 5.35* | 1.45 | |
| Invasive perennial groundcovers | 0.26 | 0.48 | 1.21 | |
| Native perennial groundcovers | 0.07 | 0.21 | 0.70 | |
| Native woody shrubs | 0.40 | 1.39 | 0.39 | |

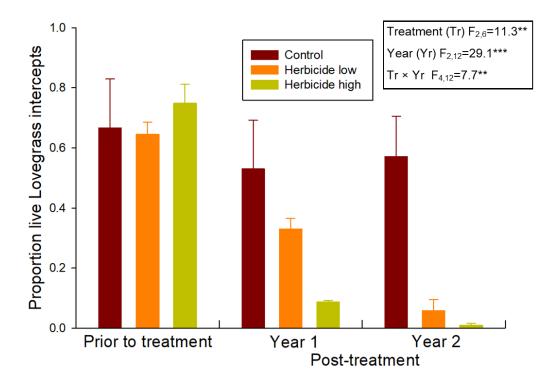
393 *** P<0.001, ** P<0.01, * P<0.05.



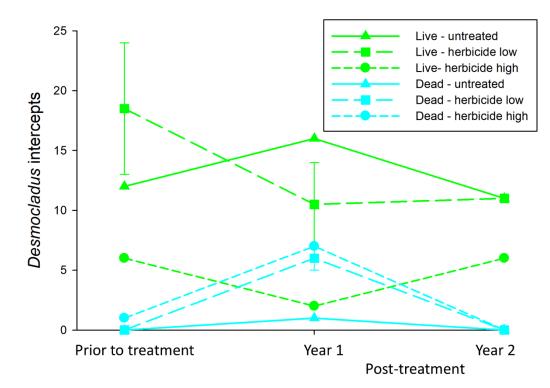








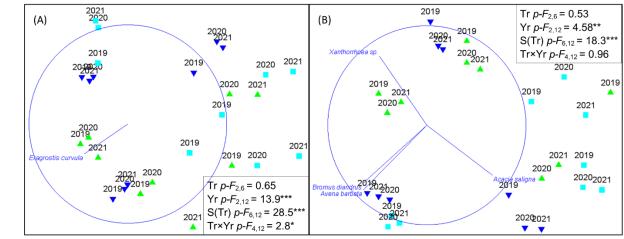
401 Fig. 2



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406 Fig. 4